INFLUENCE OF AFLATOXIN B, ON GROWTH, PHOTOSVNTHETIC OXYGEN EVOLUTION AND REGREENING OF CHLORELLA FUSCA (CHOROCOCCALES, CHLOROPHYTA)

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ABSTRACT - Treatment of the green alga Chlorelle fuzzo Shih, et Krauss with allocatin BR (AFR4) induced a variety of toxics responses including cestation of cell division and inhibition of photosynthetic oxygen evolution. Other AFB, induced metabolic disorders involved inhibition of chlorophyll symbaliss and reduction of cell contents of carbon, mitrogen, and phosphorus. Light-induced regreening of orange nitrogen-deficient cells was also inhibited.

KEY WORDS : Aflatoxin B1, Chlorella, oxygen evolution, regreening,

INTRODUCTION

AFB, is a potent hepatocarcinogen known to induce toxic effects in animats that include inhibition of DNA dependent RNA synthesis and changes in liver mitochondria (Sporn et al., 1966). In plants, AFB, has been reported to inhibit germination and root growth of soybeans (Jones et al., 1980), as well as death of floral huds (Reiss, 1969). Moreover, AFB, was shown to inflibit growth of *Chlorella prenolosas* Chick (Sultivan & Kawaa, 1972), and chlorophyll synthesis in mung leaves (Sinha & Kumari, 1960). Reviews in the literature emphasiss the meed for work on she dimore light on the effect of AFB, on the photoiss the meed for work on she horophylic hyperening experiment, and the ophylic influence of AFB, on growth, and photosynthesic oxygen evolution in Ches the faces Shih. et Krauss which through its depresenting capability provides an ideal model for examining the effects of AFB, on developing photosynthesic machinery.

MATERIALS AND METHODS

Chlorella fuzza 211-15 from the Collection of Algal Cultures (Gottingen, Germany) was grown in 12th day.night cycles in a nitrate-rich medium (Grimme & Porra, 1974). Cells were degreened at 25°C and continuous illumination of 250 µmol $m^2 s^1$ in a similar but a nitrate-sparse medium (Grimme & Porra, 1974). For representing, orange cells were harvested, washed and resus-

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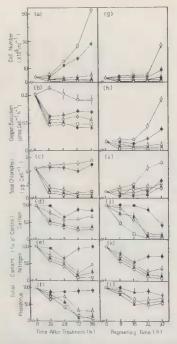


Fig. 1. - The effect of AFB₁ concentrations on growth and various physiological parameters of green (a-1) and orange mitrogen deficient (a-1) cells of Chlorella futca. (o) Control. (a) 0.1. (a) 1.0. (b) 1.0. (b) 1.0. and (□) 100,mi⁻¹ (4, ES, n = 3). Control values (pg.cell⁺) for green cells were 60 C, 5.3 N, 4.1 P, and for orange cells: 60 C, 5.4 N, 58 P.

pended in a nitrate-rich medium under continuous illumination. Cultures were sparged with fine bubbing at 25° C. AH B, in 0.03*6, dimethylsulfoxide was added to give concentrations in the range of 0.1-100 µgml⁻¹. Dimethylsulfoxide (0.03%) was also added to control cultures. Cell numbers were determined with a haemocytometer (Richert Jung, LSA) and Cidorophyll was determined after Boger (1964). Oxygene volution was measured in an oxygen electrode (Rank Brothers, UK) at 25° C and a quantum flux density of 859µmol m⁻² s⁻¹. For analysis, cells were harvested, washel and the contents of carbohydrates, nirrogen and phosphorus were determined after Dubois *et al.*, (1955), Vogel (1968) and Jackson (1960), respectively.

RESULTS

Treatment of green cells with 0.1ggmt⁻¹ resulted in 50% inhibition of growth with higher tevels causing complete inhibition (Fig. 1a). Oxygen evolution was inhibited 24h after treatment, with high levels causing a marked inhibition and a reduction in chlorophyll content (Fig. 1b and e.). Contents of carbon, nitrogen and plocsphorus of treated cells gave reduced values when expressed as percentages of their corresponding control at each measurement. These values also exhibited declining trends in a pattern that was AFB₁-concentration dependent (Fig. 1d, e., and f. respectively.)

Regreening of *Chlorella* cells resulted in a 14-fold increase in cell number which was inhibited 327-b by 0.1 graft 3.4 B, with higher levels causing complete inhibition (Fig. 1g). Regreening was complete within 24h as both oxygen evolution (Fig. 1h) and chlorophyll content (Fig. 1h) reached levels compatable to those of normal green cells (Fig. 1b, and c). Freatments resulted in further reduction of the residual oxygen evolving activity of the nitrogen-deficient (edis (Fig. 1h) as well as complete inhibition of regreening (Fig. 1h). addition, treatments resulted in reduction of cell contents of carbon, nitrogen, and phosphorus (Fig. 1), k and I, respectively).

DISCUSSION

ATB₁ caused retardation of cell division, with high levels causing complet cessation of this process in both green and orange cells. The increase in cell muther of control regreening cells indicated that each cell in the population had divided to give approximately eight diaghter cells (Fig. 1g). Cells treated with 0.1 µmt³ attained a slower rate denoting some 75° inhibition of cell division and cells treated with higher levels failed to increase in runnher. AFB, is some and cells treated with higher levels failed to increase in runnher. AFB, is DNA-dependent RAA polymerase (Dathek & Lissellon, 1963). Reduction of nitrogen contents of AFB, treated cells (Fig. Le and k) may also reflect induced inhibition of protein synthesis as has previously been reported (Tripathi & Misra, 1981).

Oxygen evolution by green cells was inhibited by treatments with AFB, Inhibited photosynthesis is presumably largely responsible for the observed reduced carbon content. In orange cells AFB, caused reduction of the residual oxygen evolving activity and their failure to regreen. Chlorophyll synthesis and development of an oxygen evolving capability during regreening of orange introgen deficient cells are considered to he two separable processes. The former being dependent on aerobic conditions, and the latter accompanies the regreening only in the light (Grimme, 1978). Inhibition of oxygen evolution in green cells at 0.1 μ gml⁻¹ occurred without reduction of chlorophyll content (Fig. 1b and c) suggesting m direct inhibitory effect of AFB₁ on photosynthesis itself, possibly at the electron transport chain level.

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